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## **XX Congresso Farmacêutico de São Paulo**

**III SIMPÓSIO “FRONTEIRAS NAS CIÊNCIAS FARMACÊUTICAS”  
10 e 11 de Outubro de 2019**



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# Zebrafish

## Sua Importância na Avaliação de Toxicidade



**Mônica Lopes Ferreira**

# Modelo experimental para pesquisa científica

Em **1981** - George Streisinger, biólogo, professor da Universidade de Oregon

Considerado o introdutor do zebrafish como modelo na pesquisa científica.

Mostrou vantagens na utilização do peixe para estudos genéticos

publicou na Nature, artigo considerado um marco na medicina translacional.





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# *Danio rerio*



***Zebrafish – Paulistinha***



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**Best matches for zebrafish:**

- [Zebrafish in Toxicology and Environmental Health.](#)  
Bambino K et al. Curr Top Dev Biol. (2017)
- [Zebrafish: an important tool for liver disease research.](#)  
Goessling W et al. Gastroenterology. (2015)
- [The developing utility of zebrafish models of neurological and neuropsychiatric disorders: A critical review.](#)  
Fontana BD et al. Exp Neurol. (2018)

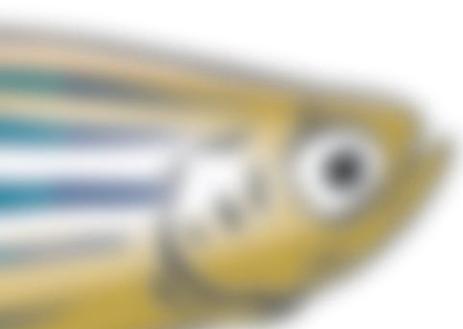
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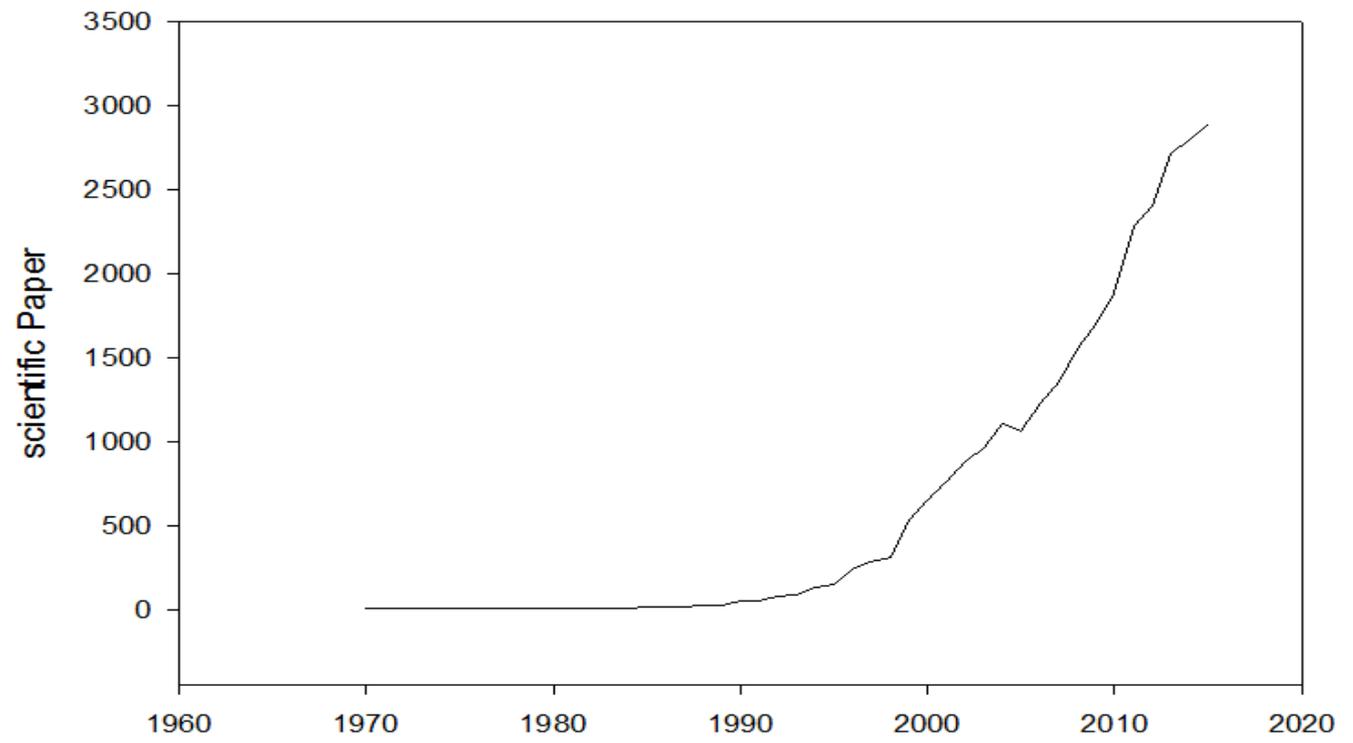
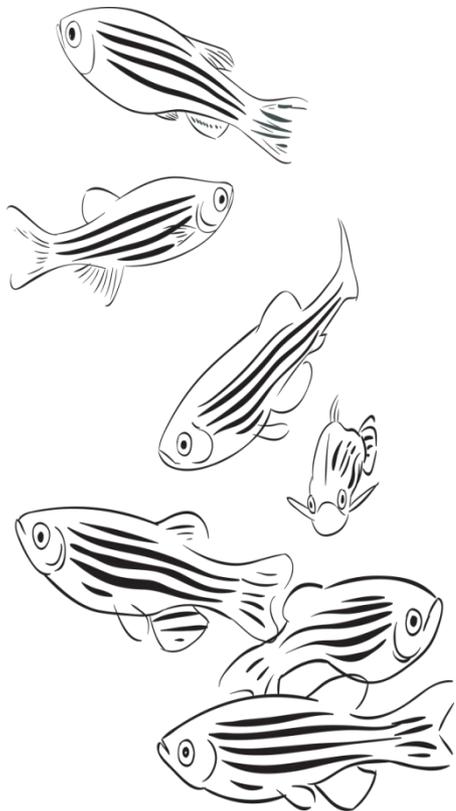
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- [Zebrafish as a Model of Neurodevelopmental Disorders.](#)  
de Abreu MS, Genario R, Giacomini ACVV, Demin KA, Lakstygala AM, Amstislavskaya TG, Fontana BD, Parker MO, Kalueff AV. Neuroscience. 2019 Aug 28; pii: S0306-4522(19)30607-4. doi: 10.1016/j.neuroscience.2019.08.034. [Epub ahead of print]  
PMID: 31472215 [Similar articles](#)
- [Isolation and Culture of Primary Embryonic Zebrafish Neural Tissue.](#)  
Patel BB, Clark KL, Kozik EM, Dash L, Kuhlman JA, Sakaguchi DS. J Neurosci Methods. 2019 Aug 28;108419. doi: 10.1016/j.jneumeth.2019.108419. [Epub ahead of print]  
PMID: 31472190 [Similar articles](#)
- [Integrated K+ channel and K+Cl- cotransporter function are required for the coordination of size and proportion during development.](#)  
Lanni JS, Peal D, Ekstrom L, Chen H, Stancliff C, Bowen ME, Mercado A, Gamba G, Kahle KT, Harris MP. Dev Biol. 2019 Aug 28; pii: S0012-1606(19)30297-0. doi: 10.1016/j.ydbio.2019.08.016. [Epub ahead of print]  
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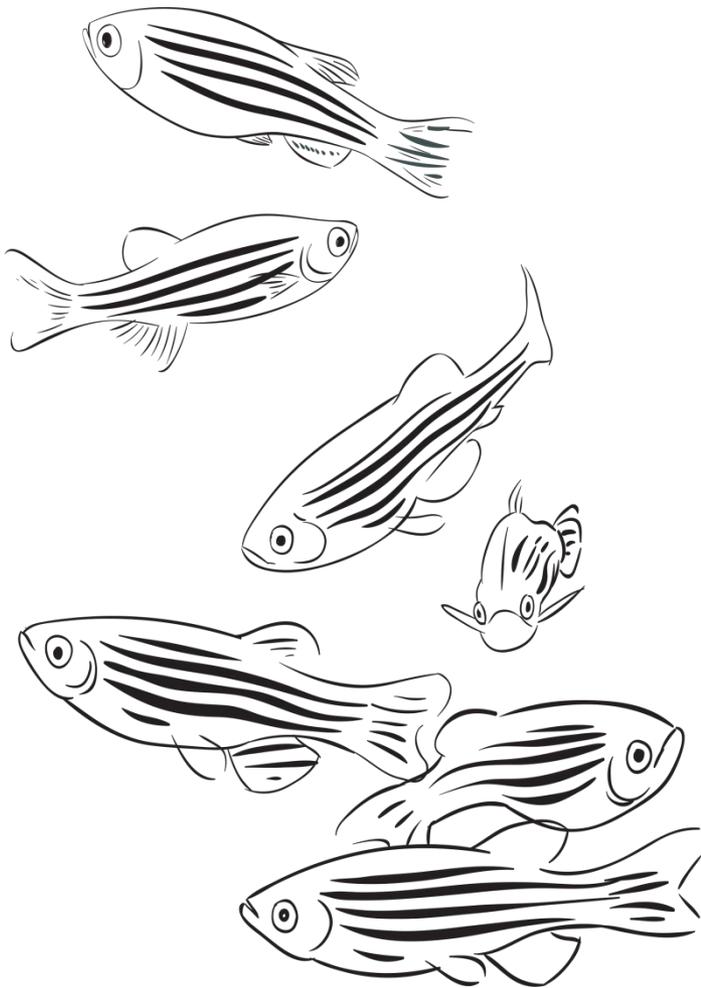


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# Zebrafish

## e Toxicidade



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[Developmental toxicity and cardiac effects of butyl benzyl phthalate in zebrafish embryos.](#)  
Sun G et al. Aquat Toxicol. (2017)

[Zebrafish in Toxicology and Environmental Health.](#)  
Bambino K et al. Curr Top Dev Biol. (2017)

[Acute and subchronic toxicity of pyraclostrobin in zebrafish \(Danio rerio\).](#)  
Zhang C et al. Chemosphere. (2017)

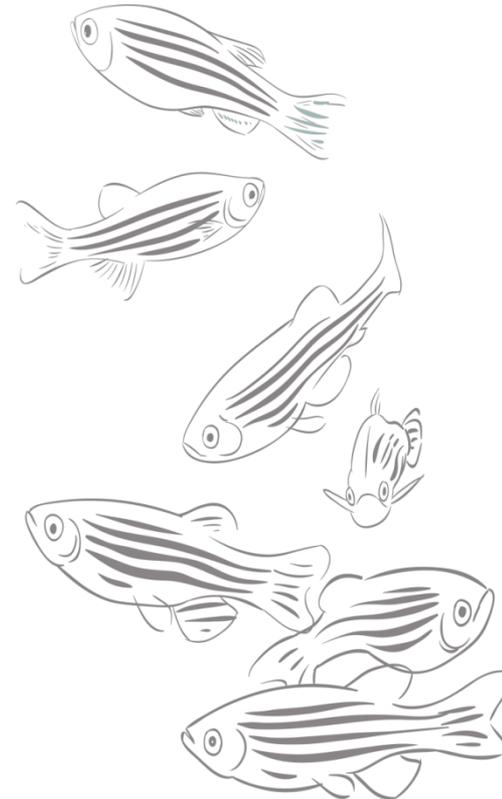
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- [Activation of the nucleotide excision repair pathway by crude oil exposure: A translational study from model organisms to the Hebei Spirit Oil Spill Cohort.](#)  
Kim Y, Jeong J, Chatterjee N, Yim UH, Kwon JH, Park MS, Choi J.  
Environ Pollut. 2019 Aug 6;254(Pt B):112997. doi: 10.1016/j.envpol.2019.112997. [Epub ahead of print]  
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- [The cardiovascular toxicity induced by high doses of gatifloxacin and ciprofloxacin in zebrafish.](#)  
Shen R, Yu Y, Lan R, Yu R, Yuan Z, Xia Z.  
Environ Pollut. 2019 Jul 9;254(Pt B):112861. doi: 10.1016/j.envpol.2019.07.029. [Epub ahead of print]  
PMID: 31454568  
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- [Hepatic proteome network data in zebrafish \(Danio rerio\) liver following dieldrin exposure.](#)  
Simmons DBD, Cowie AM, Koh J, Sherry JP, Martyniuk CJ.  
Data Brief. 2019 Aug 5;25:104351. doi: 10.1016/j.dib.2019.104351. eCollection 2019 Aug.  
PMID: 31453304  
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- [Health Impact Assessment of Sulfolone on Embryonic Development of Zebrafish \(Danio rerio\).](#)  
Shah SM, Wahba M, Yu L, Achari G, Habibi HR.  
Toxicol. 2019 Aug 23;7(3). pii: E42. doi: 10.3390/toxics7030042.  
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**Best matches for zebrafish and toxicity review:**

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Santos D et al. Aquat Toxicol. (2017)
- [Zebrafish in Toxicology and Environmental Health.](#)  
Bambino K et al. Curr Top Dev Biol. (2017)
- [The Use of the Zebrafish Model to Aid in Drug Discovery and Target Validation.](#)  
Yoganantharajah P et al. Curr Top Med Chem. (2017)

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- [Nanomaterials meet zebrafish: Toxicity evaluation and drug delivery applications.](#)
- 1. Jia HR, Zhu YX, Duan QY, Chen Z, Wu FG. J Control Release. 2019 Aug 22. pii: S0168-3659(19)30506-1. doi: 10.1016/j.jconrel.2019.08.022. [Epub ahead of print] **Review.**  
PMID: 31446084 [Similar articles](#)
- [A Critical Review of Animal Models Used in Acute Myeloid Leukemia Pathophysiology.](#)
- 2. Skayneh H, Jishi B, Hleihel R, Hamieh M, Danwiche N, Bazarbachi A, El Sabban M, El Hajj H. Genes (Basel). 2019 Aug 13;10(8). pii: E614. doi: 10.3390/genes10080614. **Review.**  
PMID: 31412687 [Free Article](#) [Similar articles](#)
- [The zebrafish embryotoxicity test \(ZET\) for nanotoxicity assessment: from morphological to molecular approach.](#)
- 3. Pereira AC, Gomes T, Ferreira Machado MR, Rocha TL. Environ Pollut. 2019 Sep;252(Pt B):1841-1853. doi: 10.1016/j.envpol.2019.06.100. Epub 2019 Jun 28. **Review.**  
PMID: 31325757 [Similar articles](#)
- [Review on the use of zebrafish embryos to study the effects of anesthetics during early development.](#)
- 4. Félix L, Coimbra AM, Valentim AM, Antunes L. Crit Rev Toxicol. 2019 Jul 17;1-14. doi: 10.1080/10408444.2019.1617236. [Epub ahead of print]  
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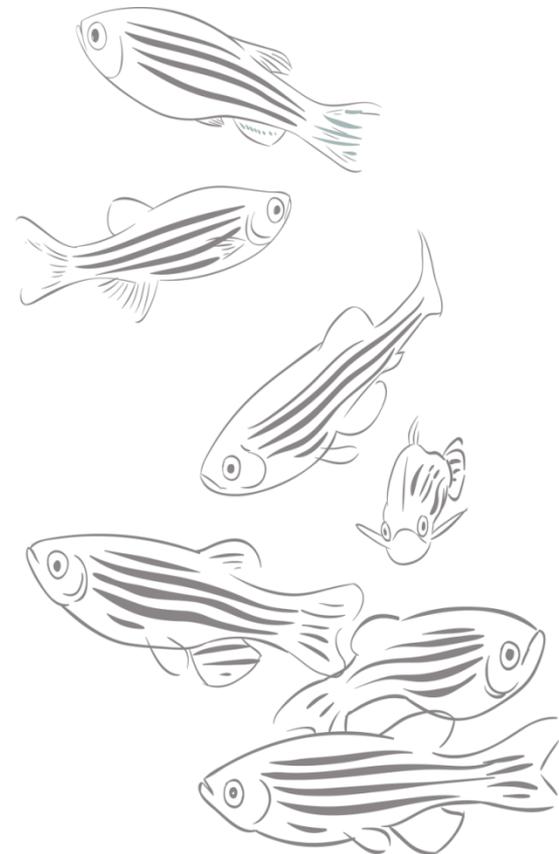
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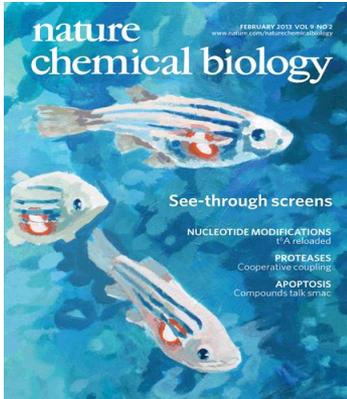
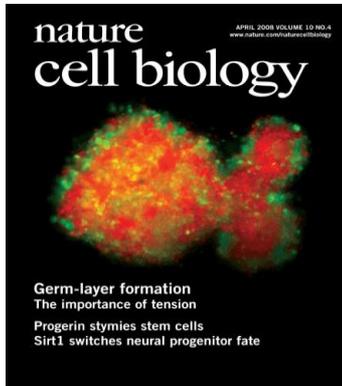
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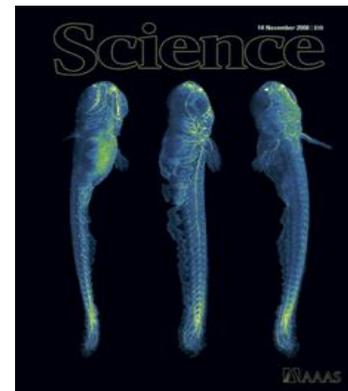
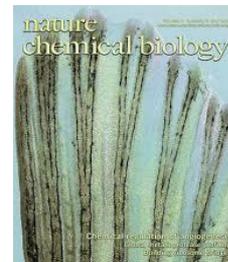
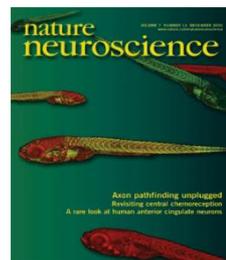
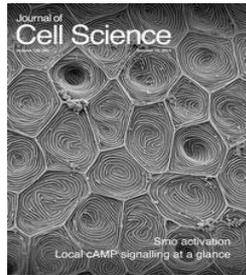
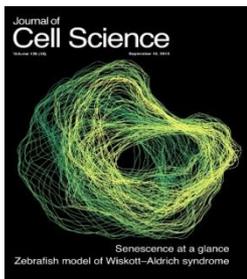




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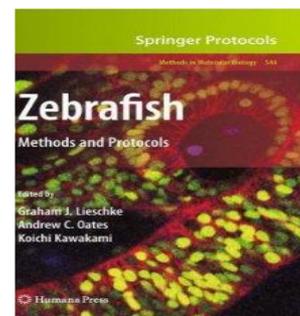
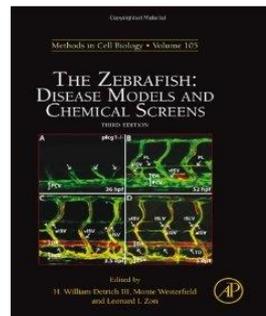
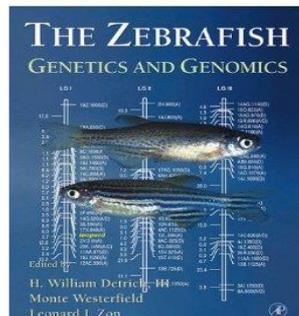
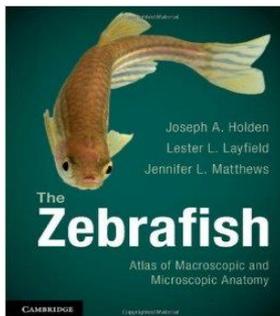
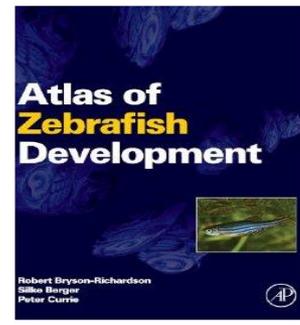
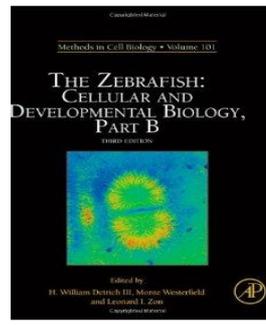
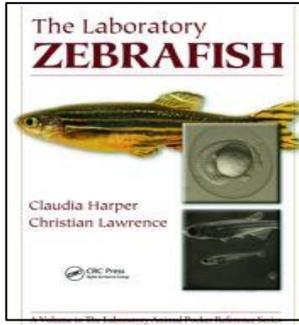
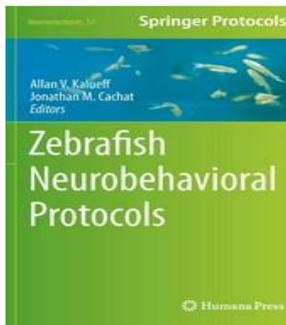
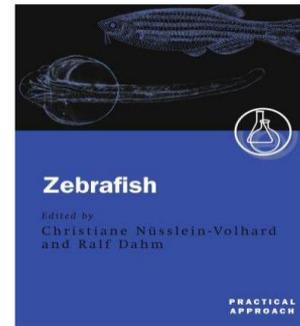
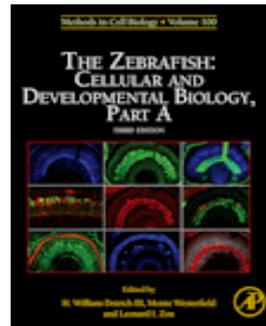
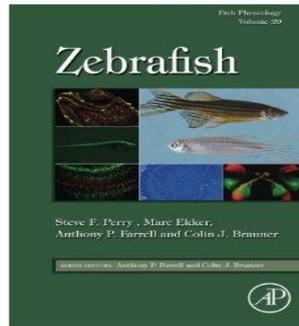
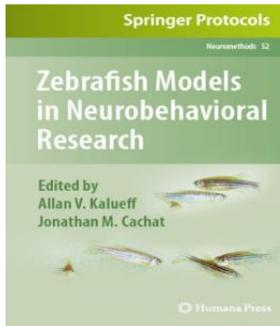


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# Importante



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Vania Gomes de Moura Mattaraia  
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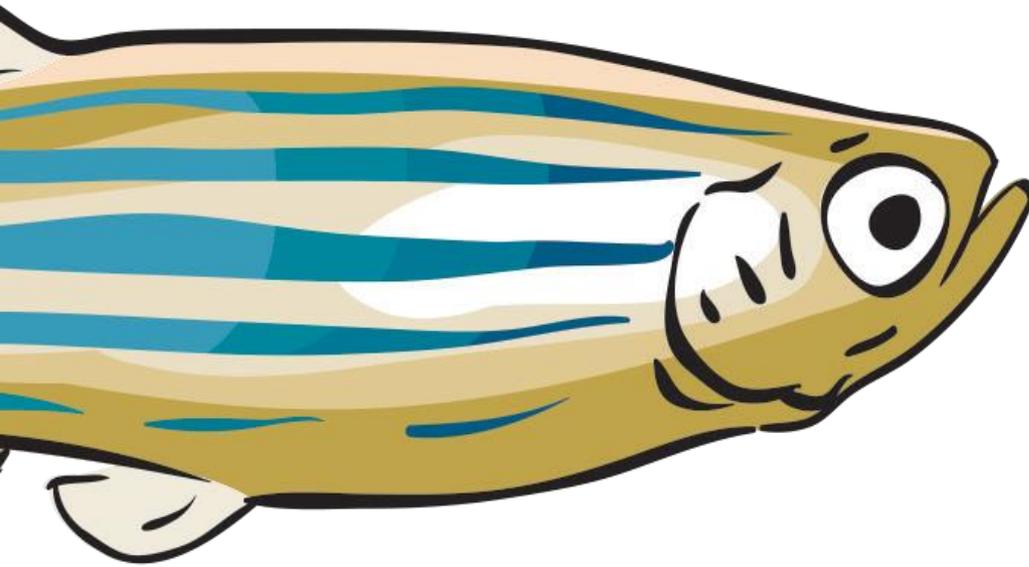
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**O uso do teleósteo *Danio rerio* como  
organismos modelo em estudo de  
sanidade aquícola**

Cintia Badaró-Pedroso  
Mônica Lopes-Ferreira  
Cláudia Marins Ferreira  
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Vantagens como modelo  
experimental



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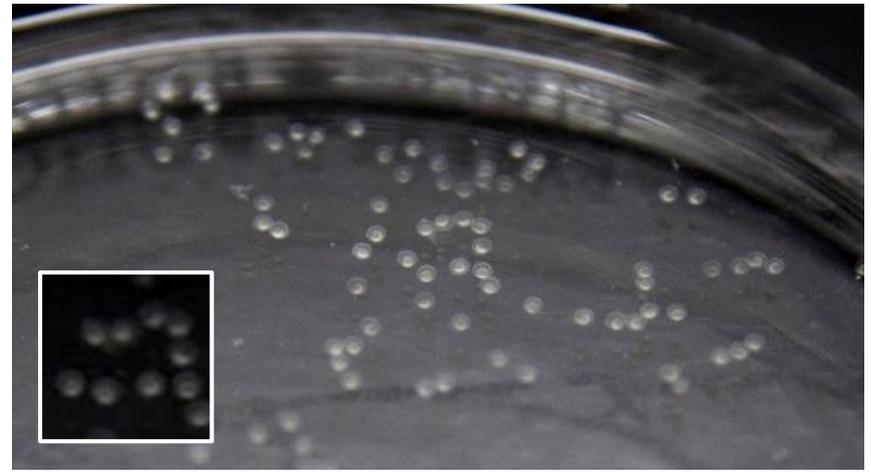
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## **Zebrafish: vantagens como modelo experimental**

1. Alta taxa reprodutiva
  2. Embriões transparentes
  3. Rápido desenvolvimento
  4. Pequeno porte
  5. Uso na experimentação de todos os estágios de vida
  6. Fácil manipulação
  7. Econômicos para criação
  8. Genoma sequenciado
  9. Métodos e estratégias de avaliação genética e embriológica são aplicados com facilidade ao modelo
  10. Importante número de mutantes
-

## Zebrafish: vantagens como modelo experimental

### 1. Alta taxa reprodutiva: 100 a 200 embriões/dia



## Zebrafish: vantagens como modelo experimental

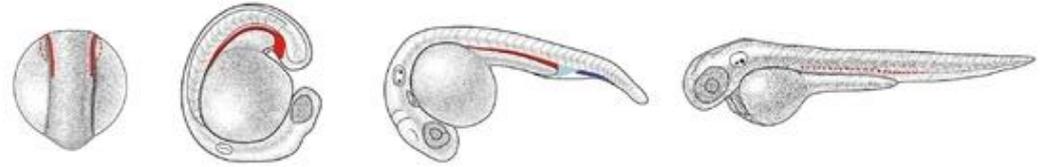
### 2. Embriões transparentes



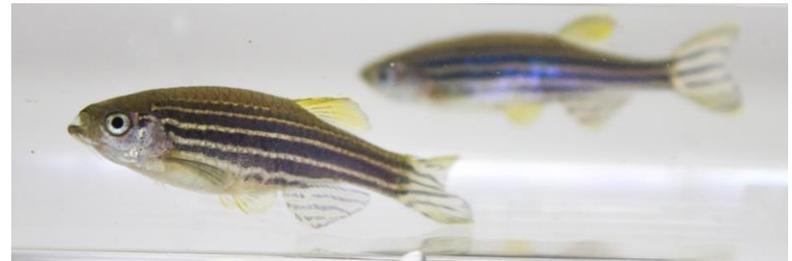
## Zebrafish: vantagens como modelo experimental

### 3. Rápido desenvolvimento

48 a 72 horas: ovo → larva



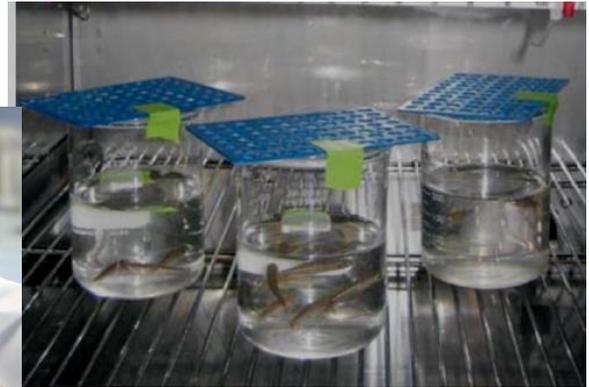
### 4. Pequeno porte: 3 a 5 cm quando adulto



# Zebrafish: vantagens como modelo experimental

## 6. Fácil manipulação

a) imersão do animal em uma solução com o agente a ser investigado



b) administração de substâncias pode ser feita por via oral, intramuscular, intravenosa e intraperitoneal



## Zebrafish: vantagens como modelo experimental

### 7. Econômicos para criação (recursos e objetivos)

	 ZEBRAFISH	 CAMUNDONGO
MANUTENÇÃO DIÁRIA	R\$ 0,60	R\$ 8,00

FONTE JOSÉ XAVIER NETO / LNBIO, MONICA RYFF VIANNA / PUC-RS E DENIS ROSEMBERG / UNCHAPECÓ



## Zebrafish: vantagens como modelo experimental

8. Genoma sequenciado: 70 % de seus 26 mil genes são semelhantes aos genes humanos



**70%** de seus 26 mil genes são semelhantes aos genes humanos quando adulto



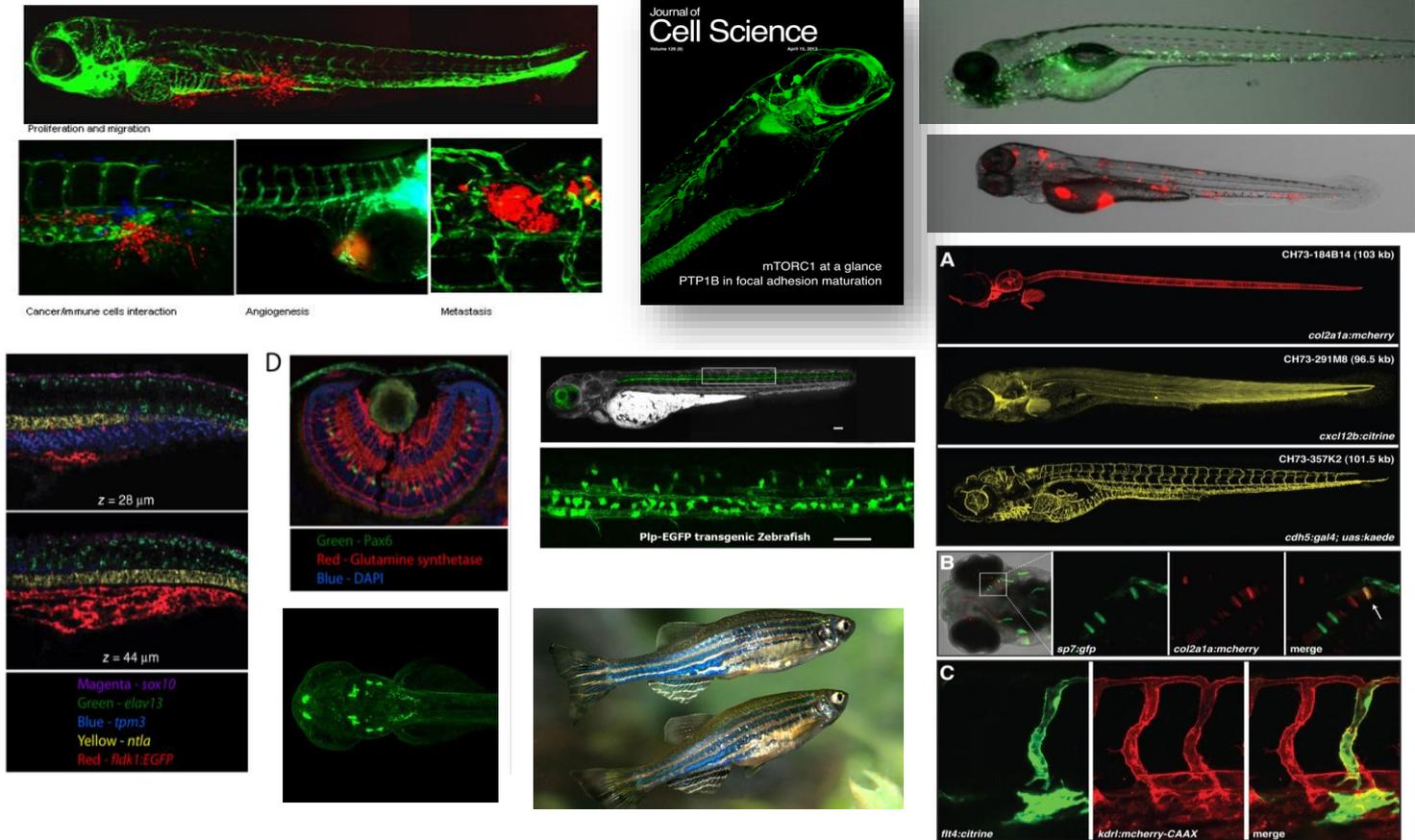
e quando avaliados somente genes envolvidos com doenças esta homologia sobre para **84%**



# Zebrafish: vantagens como modelo experimental

9. Métodos e estratégias de avaliação genética e embriológica são aplicados com facilidade ao modelo

10. O desenvolvimento de técnicas especiais de clonagem, mutagênese e transgênese permite a identificação de um número importante de mutantes



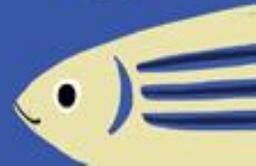
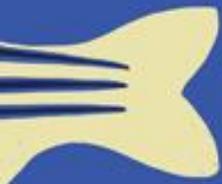
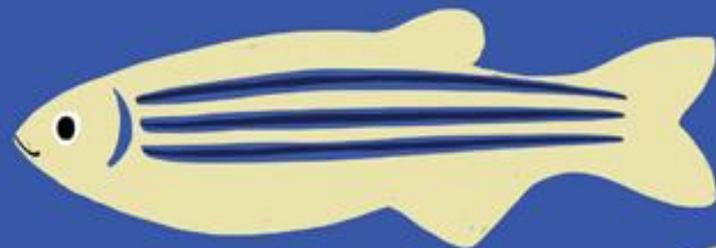
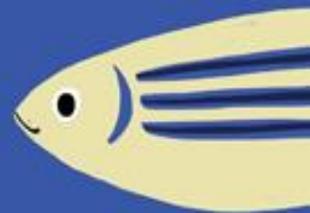


# PLATAFORMA ZEBRAFISH

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# Plataforma Zebrafish



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# Plataforma Zebrafish





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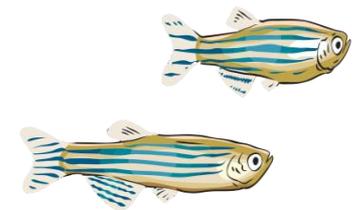
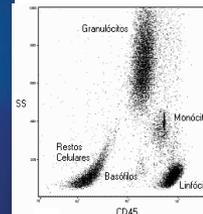
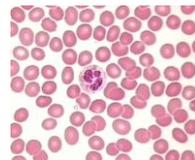
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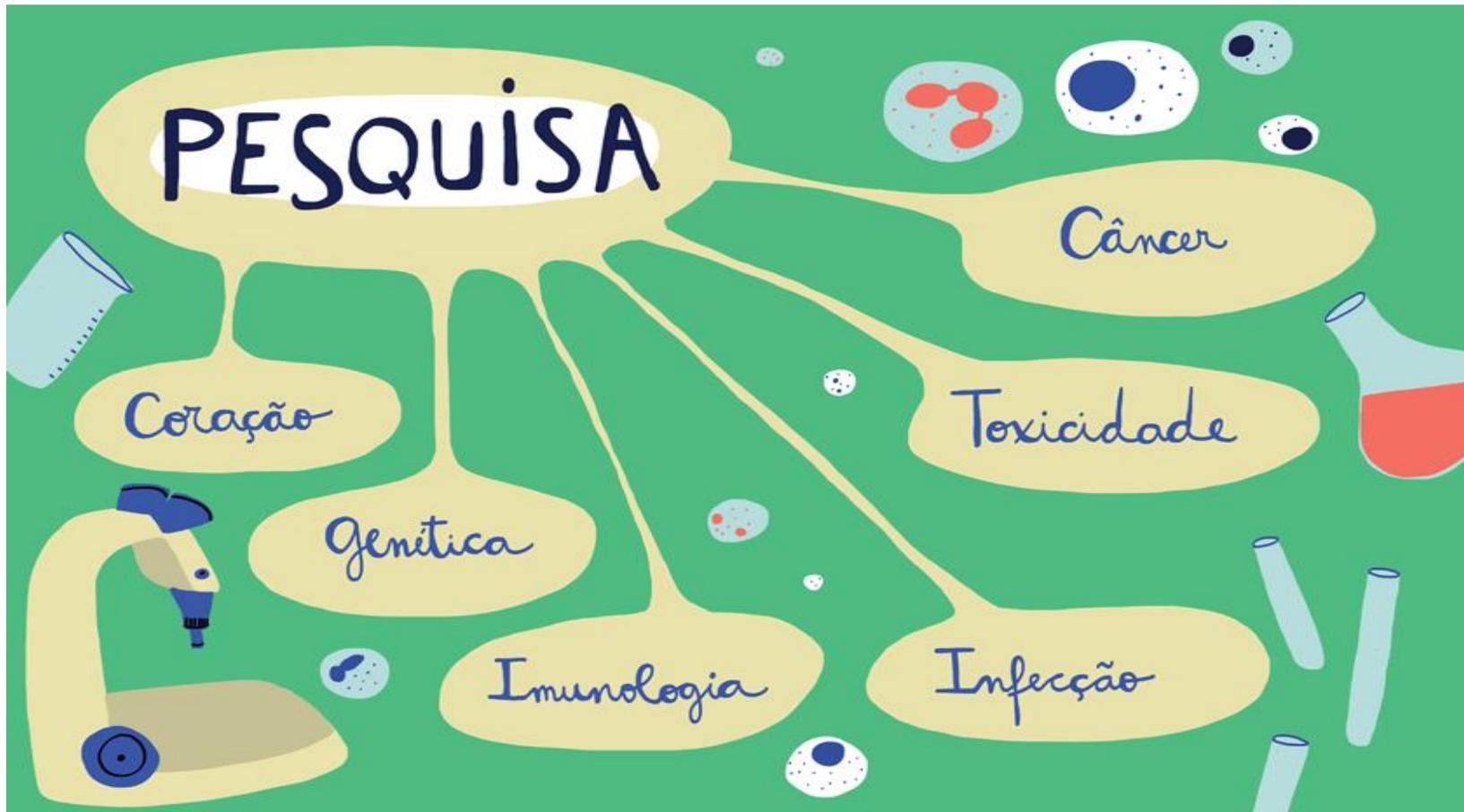
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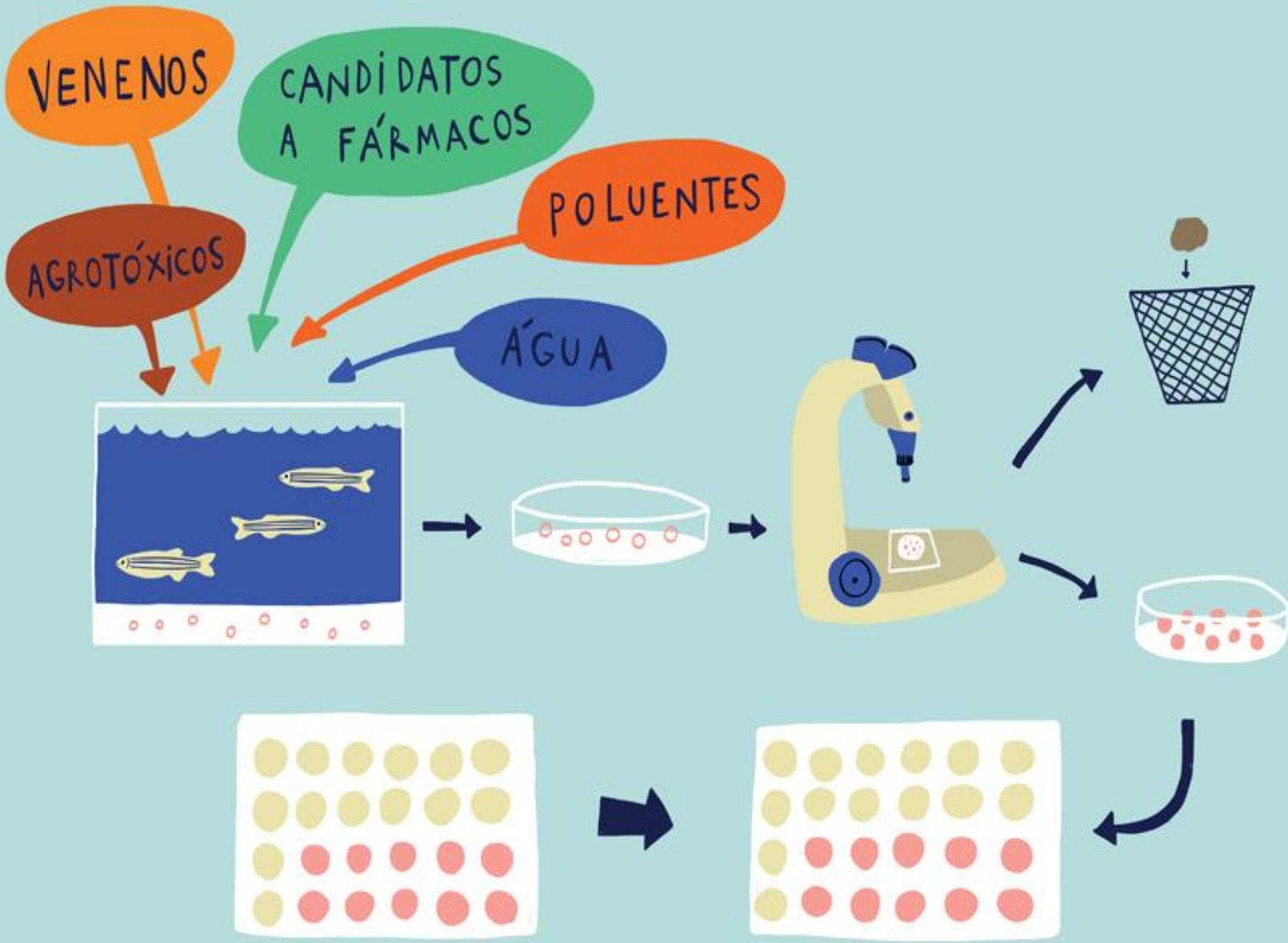
# Plataforma Zebrafish





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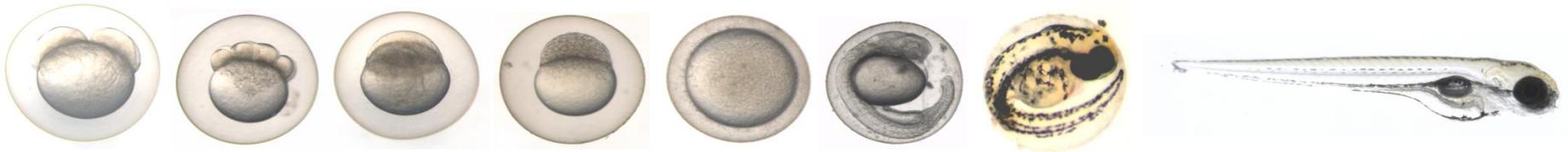
# Toxicidade

Teste Toxicidade em embriões de peixes

“Zebrafish Embryotoxicity Test” - ZET

*Danio rerio*

OECD 236

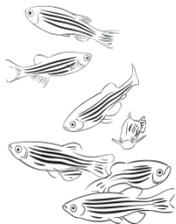


## Características Importantes para o Teste ZET (*Danio rerio*)

- \* Fertilização externa
- \* Baixo custo
- \* Necessidade de pequenas quantidades de moléculas
- \* Tempo de exposição reduzido
- \* Embriões e larvas transparentes, o que facilita a visualização de seu desenvolvimento
- \* Ciclo de vida curto
- \* Rápido desenvolvimento embrionário rápido
- \* Descoberta rápida de fenótipo
- \* Rastreabilidade genética
- \* Eticamente aceitável
- \* Grande sensibilidade quando exposto a produtos químicos por ser capaz de absorver de forma rápida os compostos que são diretamente adicionados na água e acumulá-los em diferentes tecidos (principalmente no Sistema Nervoso Central)
- \* Permite a avaliação de respostas crônicas, teratogenicidade, cardiotoxicidade, genotoxicidade, distúrbios musculares e ósseos
- \* Toxicidade neurocomportamental, toxicidade específica de órgãos (ex. hepatotoxicidade e nefrotoxicidade), toxicidade reprodutiva, ruptura endócrina, estresse oxidativo e avaliação de risco ambiental



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Adopted:  
26 July 2013

## OECD GUIDELINES FOR THE TESTING OF CHEMICALS

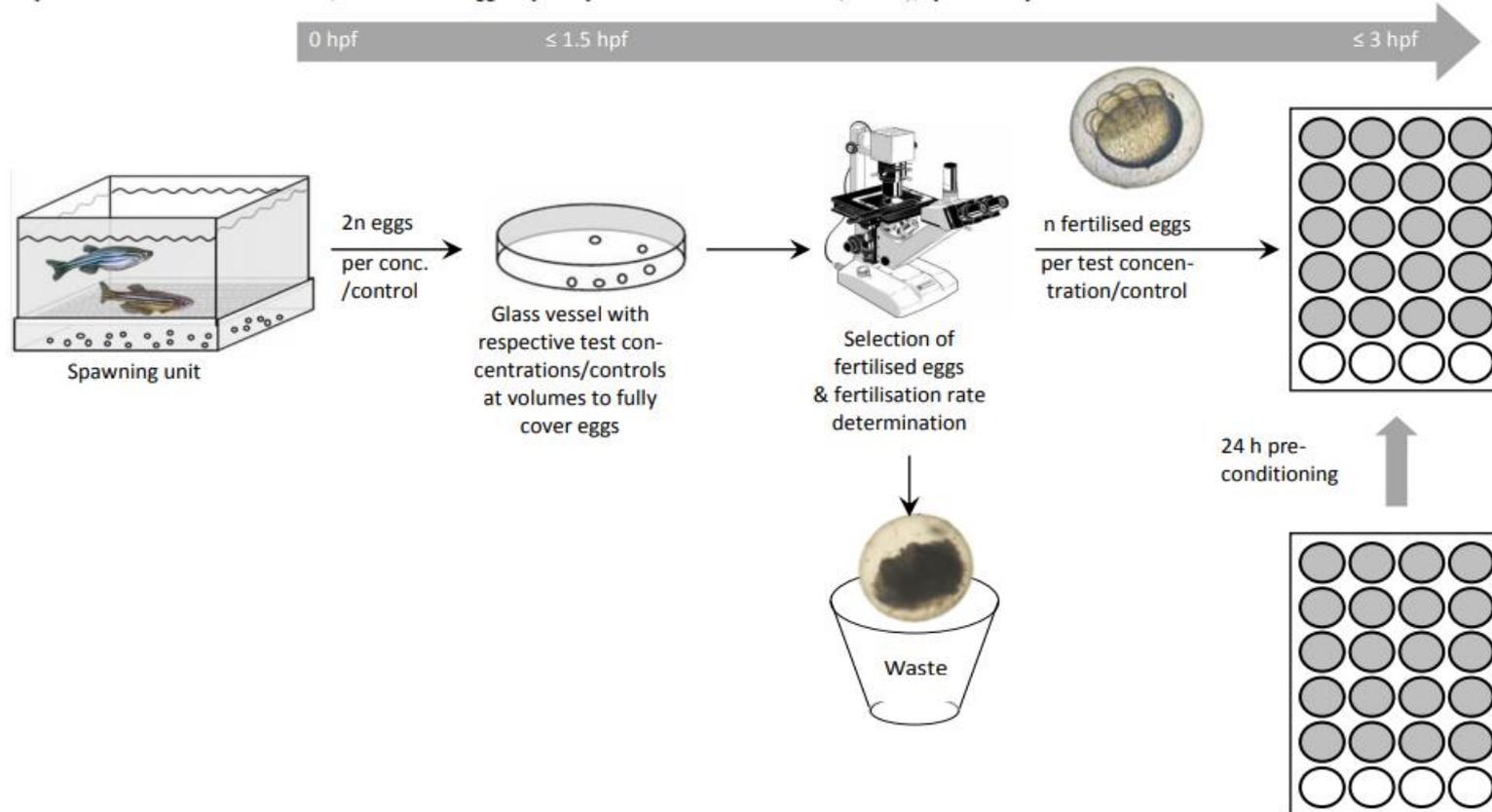
### PRINCIPLE OF THE TEST

3. Newly fertilised zebrafish eggs are exposed to the test chemical for a period of 96 hrs. Every 24 hrs, up to four apical observations are recorded as indicators of lethality (6): (i) coagulation of fertilised eggs, (ii) lack of somite formation, (iii) lack of detachment of the tail-bud from the yolk sac, and (iv) lack of heartbeat. At the end of the exposure period, acute toxicity is determined based on a positive outcome in any of the four apical observations recorded, and the LC<sub>50</sub> is calculated.

Embriões de zebrafish recém-fertilizados (no estágio da blástula  $\leq 3$  hpf) são expostos a produtos químicos em teste **por um período de 96 horas**.

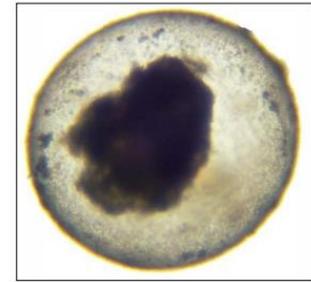
**A cada 24 horas**, até quatro observações são registradas **como indicadores de letalidade**: (1) coagulação de ovos fertilizados, (2) falta de formação de somito, (3) falta de descolamento da cauda e (4) ausência de batimento cardíaco.

Fig. 2: Scheme of the zebrafish embryo acute toxicity test procedure (from left to right): production of eggs, collection of the eggs, pre-exposure immediately after fertilisation in glass vessels, selection of fertilised eggs with an inverted microscope or binocular and distribution of fertilised eggs into 24-well plates prepared with the respective test concentrations/controls, n = number of eggs required per test concentration/control (here 20), hpf = hours post-fertilisation.



**Table 1.** Apical observations of acute toxicity in zebrafish embryos 24 - 96 hrs post fertilisation.

	Exposure times			
	24 hrs	48 hrs	72 hrs	96 hrs
Coagulated embryos	+	+	+	+
Lack of somite formation	+	+	+	+
Non-detachment of the tail	+	+	+	+
Lack of heartbeat		+	+	+



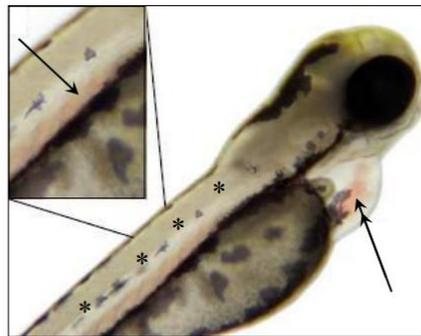
**Fig. 1: Coagulation of the embryo:** Under bright field illumination, coagulated zebrafish embryos show a variety of intransparent inclusions.

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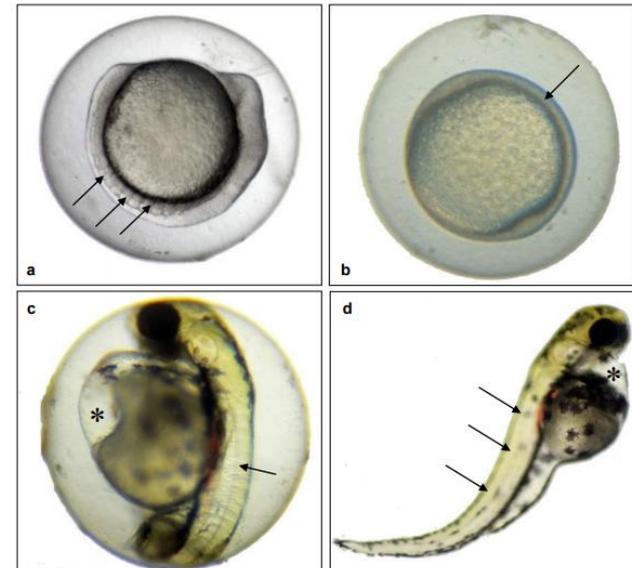
**Fig. 3: Non-detachment of the tail bud** in lateral view (a: →; 96 hrs old zebrafish embryo). Note also the lack of the eye bud (\*).



**Fig. 4: Lack of heartbeat** is, by definition, difficult to illustrate in a micrograph. Lack of heartbeat is indicated by non-convulsion of the heart (double arrow). Immobility of blood cells in, e.g., the aorta abdominalis (→ in insert) is not an indicator for lack of heartbeat. Note also the lack of somite formation in this embryo (\*, homogenous rather than segmental appearance of muscular tissues). The observation time to record an absence of heartbeat should be at least of one minute with a minimum magnification of 80×.

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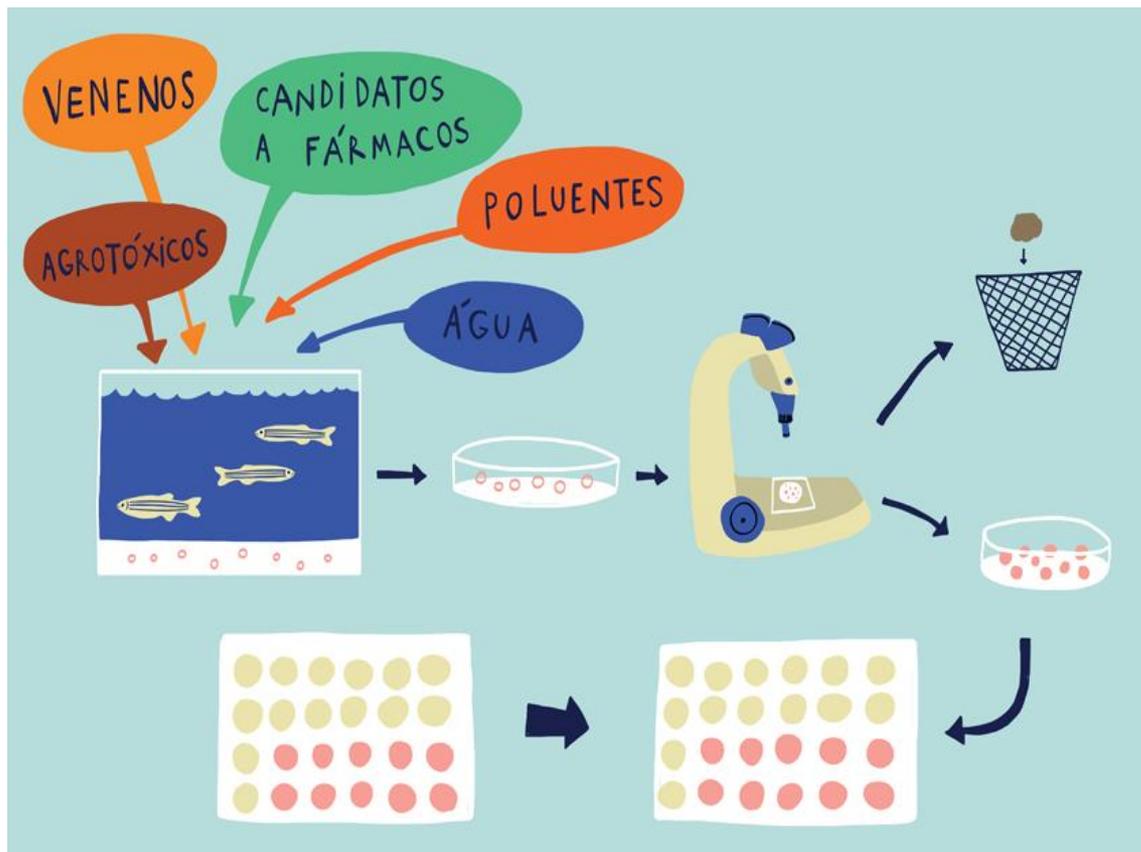


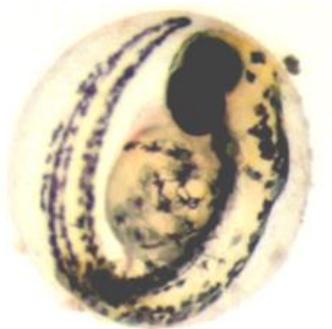
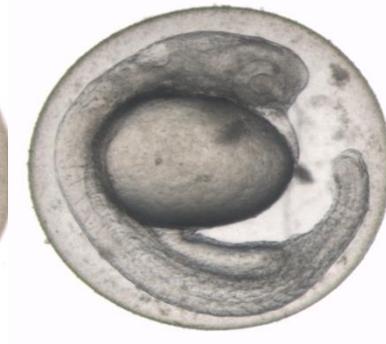
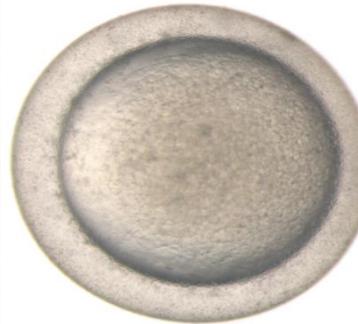
**Fig. 2: Lack of somite formation:** Although retarded in development by approx. 10 hrs, the 24 hrs old zebrafish embryo in (a) shows well-developed somites (→), whereas the embryo in (b) does not show any sign of somite formation (→). Although showing a pronounced yolk sac oedema (\*), the 48 hrs old zebrafish embryo in (c) shows distinct formation of somites (→), whereas the 96 hrs old zebrafish embryo depicted in (d) does not show any sign of somite formation (→). Note also the spinal curvature (scoliosis) and the pericardial oedema (\*) in the embryo shown in (d).



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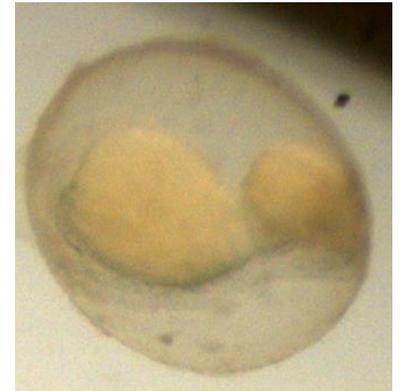
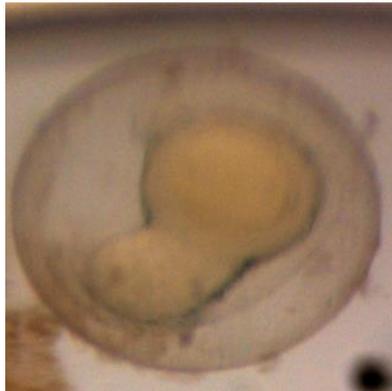
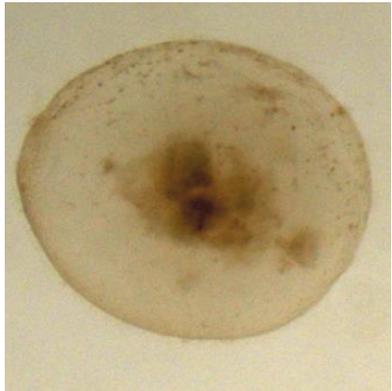
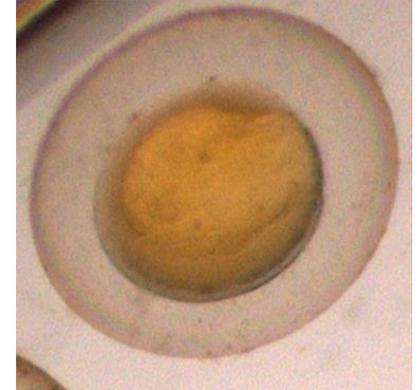
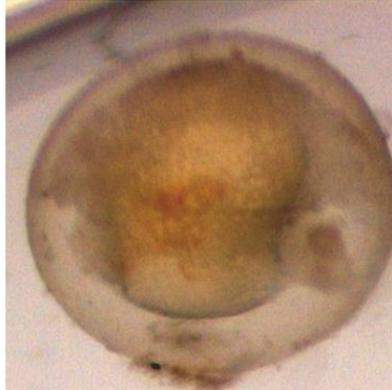
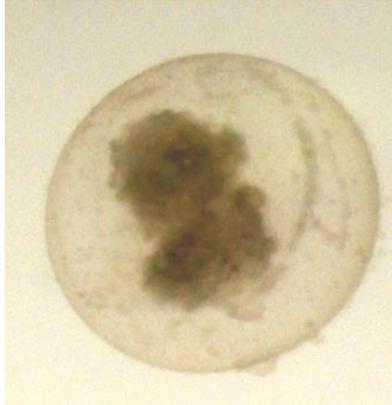






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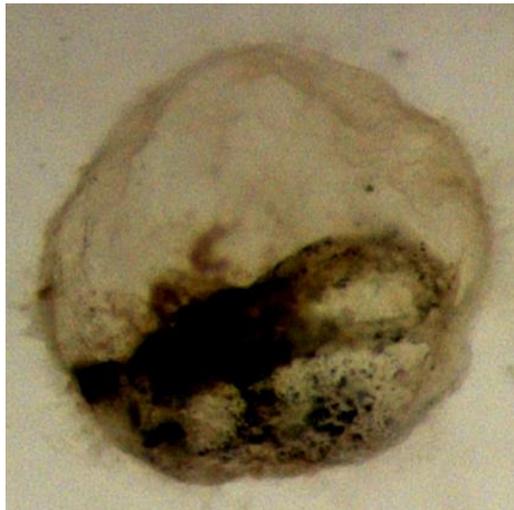
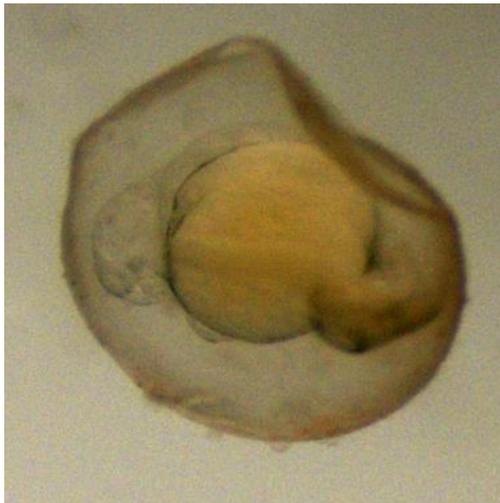
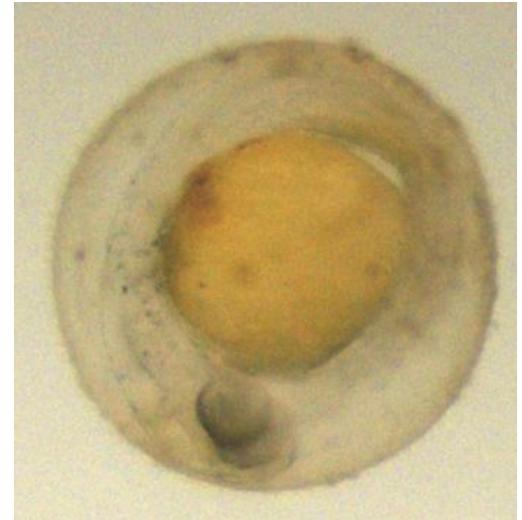
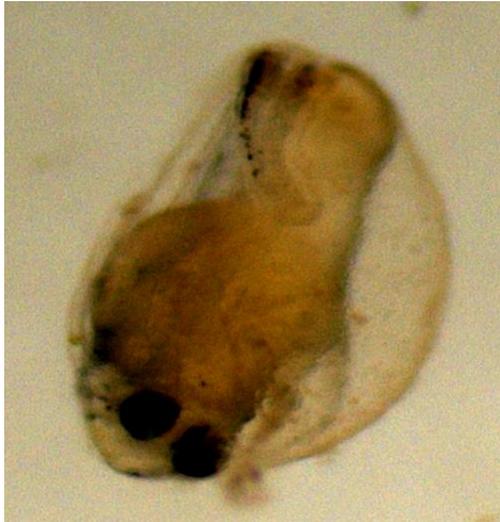
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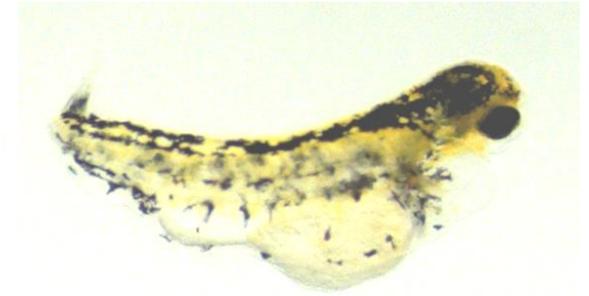
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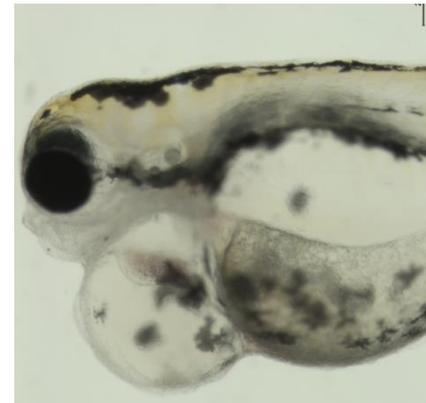
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*Muito obrigada!!*



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